Development of Stable, Orally Bioavailable, Small-Molecule **Positive Modulators of** HGF/MET Signaling for the Treatment of **Cognitive Impairment**

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CONCLUSIONS

The design and synthesis of novel HGF/MET positive modulators produced a series of active small molecules with favorable physiochemical properties

Several compounds, including the two example compounds (compounds 2 and 6), were orally bioavailable and distributed efficiently to the brain

Oral administration of compound 2 or 6 in a chemically induced model of spatial memory deficit significantly restored cognitive performance

KEY TAKEAWAY

Novel, orally bioavailable small-molecule HGF/MET positive modulators were distributed efficiently to the brain and rescued spatial memory deficits; based on these promising preclinical results, these compounds are being developed as potential therapeutic agents for Alzheimer's disease and other neurological disorders



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and inhibit inflammation

HGF/MET signaling promotes neuroprotective, neurotrophic, and anti-inflammatory mechanisms¹⁻⁴

- of the resulting compound series
- are reported

To develop novel, small-molecule positive modulators of the HGF/MET neurotrophic system that are orally bioavailable, can cross the blood-brain barrier, and improve cognitive performance in a rodent model of spatial memory deficit



INTRODUCTION

Figure 1. Positive modulators of the HGF/MET pathway may promote regenerative processes



- MET expression is reduced in neurodegenerative disorders such as Alzheimer's disease⁵
- Based on 3-dimensional conformation modeling of first-generation HGF/MET modulators, a series of cyclized compounds were generated, with a focus on increasing oral bioavailability while reducing both molecular weight and rotatable bonds
- A screening funnel was developed to evaluate the in vitro activity, physiochemical properties, oral brain distribution, and in vivo activity
- Two example compounds that are orally bioavailable, brain penetrant, and capable of reversing chemically induced spatial memory deficits

OBJECTIVE

METHODS

Figure 2. A series of cyclized compounds based on modeling of first-generation HGF/MET positive modulators were processed through a screening funnel to identify compounds with sufficient activity, stability, bioavailability, and in vivo activity to proceed to clinical development

(A) Each stage of the screening process proceeded stepwise to identify activity, stability, physiochemical properties, PK, and in vivo activity. (**B**) Efficacy was assessed through a behavioral assay using the MWM.

In Vitro

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- MET activation with various
- Cell scatterin
- molecules and
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In Vivo

- PK was asses the AUC of P
- Brain distribu
- After 15 mi target tissu Test compo
- Predicted bioavailab By use of the
- Behaviora
- The time to

Figure 3. Several compounds were identified for further development based on in vitro activity and favorable physiochemical properties

Compo	und R1	R2	R3	pMET ELISA	Cell scattering	Solubility	Stability	Permeability
2	O_{R^1} F	R ²	CH3					
3	R ¹ OH	\mathbb{R}^2	CH3					
6	R^1 F	R ²	CH3					
8	O R1 OH	R ²	CH3					
12	CI F F $O_{R^1} F$	R ²	CH3					
30		R ²	CH3					
32	O_{R^1} F	0 — ^{R²}	CH3					
35	O_{R^1} F	$H_2 N $	CH3					
49	R^1 F	R ²	R ³ OH					
50	R^1 F	R ²	R ³ NH ₂					

	Fi
es of chemicals were synthesized in solution-phase chemistry, and molecules were characterized by HPLC and NMR	
mical properties were calculated by use of DataWarrior (OpenMolecules)	
on was assessed using a pMET sandwich ELISA kit (Cell Signaling Technologies) in HEK-293 cells that were incubated for 15 minutes small molecules and a subthreshold dose of HGF (1 ng/mL)	
ng was assayed by imaging MDCK cells – labeled with a nonspecific membrane stain (WGA-488) – treated for 24 hours with small Ind a subthreshold dose of HGF (5 ng/mL)	
ability, and permeability were also assessed (Supplemental Methods, QR code)	
ssed in Sprague Dawley rats that received small molecules (IV or PO); bioavailability was assessed by comparing YK curves	
ution was assessed after IV administration of novel small molecules	
inutes, animals were anesthetized by use of isoflurane and perfused with PBS before dissection and homogenization of Je	
ounds were quantified by detection via LC-MS/MS	
unbound brain exposure was calculated under consideration of dose normalization, nonspecific protein binding, oral ility, and brain distribution	
e MWM, spatial memory performance was evaluated in rats	
I data were assessed 5 times on each of 8 consecutive days	
o find the submerged escape platform (escape latency) was recorded for each trial	

60 minutes before testing, rats received test compounds (indicated dose, PO)

20 minutes before testing, rats received a cholinergic transmission inhibitor (scopolamine, 3 mg/kg IP) to induce an amnesic state

RESULTS

R region key



P* = 0.033; *P* = 0.004.

References 1. Ebens A et al. Neuron. 1996;17:1157-1172. 2. Maina F, Klein R. Nat Neurosci. 1999;2:213-217. 3. Shang J et al. J Neurosci Res. 2011;89:86-95. **4.** Nakamura T, Mizuno S. Proc Jpn Acad Ser B Phys Biol Sci. 2010;86:588-610. **5.** Hamasaki H et al. Neuropathology. 2014;34:284-290.

Acceptable Unacceptable Not tested

Of 74 compounds, 10 are highlighted to exemplify the various R regions (R1, R2, and R3) explored in this set of cyclized compounds. Each compound's performance in a series of assays designed to assess activity and druglike properties was used to determine whether they were acceptable (turquoise) or unacceptable (grey) for ongoing development.^a White indicates that the compound was not tested in the indicated assay. Based or these results, a series of compounds were selected for further development. Here we present example data related to compound 2 and compound 6 (indicated by orange and blue boxes). ^aDevelopment thresholds are as follows: pMET ELISA, statistically significant augmentation of pMET above HGF 1 ng/mL alone; cell scattering, statistically significant augmentation of colony-scattering behavior above HGF 5 ng/mL alone; solubility, aqueous solubility ≥300 uM; stability, >50% compound remaining after 4 hours in the following solutions: simulated gastric fluid, simulated intestinal fluid, rat plasma, and human plasma; permeability, greater than 2×10^{-6} cm/s as measured in the parallel artificial membrane permeability assay.

Figure 4. Tested compounds were orally bioavailable



Plasma concentration of (A) compound 2 and (B) compound 6 over time after PO or IV dosing.

Data are mean \pm SEM (n = 8 animals).

Example compounds (compound 2 and compound 6) were rapidly distributed and cleared from the plasma when administered IV; both compounds exhibited slower distribution with oral administration. Other tested compounds (not shown) also had similar bioavailability results

Table 1. Modeling of unbound concentration of compounds in the brain suggested that the cyclized compounds can achieve therapeutic doses with oral dosing

	Compound 2 Mean ± SEM, nM	Compound 6 Mean ± SEM, nM		
Whole brain	8.40 ± 1.43	1.02 ± 0.22		
Hippocampus	7.26 ± 0.98	0.90 ± 0.19		
Cerebellum	6.78 ± 2.19	1.05 ± 0.25		
Cortex	9.10 ± 1.66	1.20 ± 0.22		

Calculated brain exposure after 1 mg/kg dose PO.

Compound 2 and compound 6 both distributed to all evaluated regions of the brain. Dose modeling indicated that lower doses of compound 2 will likely be needed to reach neuroactive exposures

Figure 5. Novel compounds restored spatial memory performance in the MWM



Both example compounds demonstrate rescue of scopolamine-induced spatial memory deficits, with (A) compound 2 and (B) compound 6 showing significant restoration of spatial memory performance (2-way ANOVA). Data are mean ± SEM (n = 8 animals).

s ANOVA, analysis of variance; AUC, area under the curve; ELISA, enzyme-linked immunosorbent assay; HEK-293, human embryonic kidney 293; HGF, hepatocyte growth factor; HPLC, high-performance liquid chromatography; IP, intraperitoneally; IV, intravenously; LC-MS/MS, liquid chromatography with tandem mass spectrometry; MDCK, Madin-Darby canine kidney; MWM, Morris water maze; NMDA, N-methyl-D-aspartate; NMR, nuclear magnetic resonance; P, phosphorylation; PBS, phosphate-buffered saline; PK, pharmacokinetics; **PKC**, protein kinase C; **pMET**, phosphorylation of MET; **PO**, orally; **SEM**, standard error of the mean.